

Piptamine, a New Antibiotic Produced by *Piptoporus betulinus* Lu 9-1

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In the course of our screening for antimicrobial metabolites of basidiomycetes we isolated piptamine **1** from a submerged culture of the mushroom *Piptoporus betulinus* Lu 9-1. Here we report the production, isolation, structure elucidation and biological activity of **1**.

The strain was isolated from a forested district near Neustadt/Orla (Thuringia, Germany) and deposited in the strain collection of the Hans-Knöll-Institute of Natural Products, Jena, Germany. A malt agar slant culture of *Piptoporus betulinus* Lu-9-1 was used to inoculate 500 ml Erlenmeyer flasks containing 100 ml of a seed medium composed of glucose 1%, malt extract 2%, soya bean meal 0.5%, yeast extract 0.1%, KH_2PO_4 0.1%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.05%, $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ 0.0008%, CaCO_3 0.1%, pH 5.0–5.3 (before sterilization). Cultivation occurred at 23°C and 110 rpm on a rotary shaker for 21 days.

Thereafter, 10 liters of the broth was extracted twice with 3 liter portions of ethyl acetate. The combined extract was dried and evaporated. The oily residue was subjected to column chromatography on Sephadex LH-20 (MeOH). Antibacterial fractions were detected by agar diffusion assay with *Bacillus subtilis* ATCC 6633 as test organism, pooled and evaporated to dryness. Final purification was achieved by column chromatography on silica gel 60 (0.063–0.1 mm) eluting with CHCl_3 , $\text{CHCl}_3/\text{MeOH}$ (9:1), $\text{CHCl}_3/\text{MeOH}$ (8:2), and $\text{CHCl}_3/\text{MeOH}$ (7:3). The bioactive fractions were evaporated to yield 35 mg of **1** as waxy solid. The R_f on TLC (silica gel aluminium sheets, $\text{CHCl}_3/\text{MeOH}$; 9:1 (v/v)) was 0.1.

The molecular weight of piptamine (**1**) and the chemical formula ($\text{C}_{23}\text{H}_{41}\text{N}$) were readily determined by HRESI-MS (Finnigan MAT 95 XL): m/z 332.3307 ($[\text{M}+\text{H}]^+$ 100%, calcd. 332.3298). Daughter ions were generated by CID-MS/MS of m/z 332.3307 (ESI-triple quadrupol instrument Quattro, VG Biotech, Altrincham, England). Thus diagnostic fragments with m/z 240.5 and m/z 90.7 were attributable to the methylpentadecylamino moiety and the

benzyl group, respectively. In addition to m/z 332 ($[\text{M}+\text{H}]^+$) the ESI-MS of several products of other fermentations displayed m/z 304.3 ($[\text{M}+\text{H}]^+$, up to 30%) and m/z 360.4 ($[\text{M}+\text{H}]^+$, up to 15%), suggesting the occasional presence of chromatographically inseparable homologues. In accord with this contention CID-MS/MS of m/z 304.3 and m/z 360.4 furnished m/z 212.2 and m/z 90.7, respectively, and m/z 268.6 and m/z 90.6 as diagnostic fragments, suggesting that both minor components were distinguishable from **1** by a variable length of the side chain.

The structure of **1** was settled conclusively on the basis of one- and two-dimensional ^1H and ^{13}C NMR measurements (COSY, DEPT, HSQC, HMBC; Bruker Avance DRX 500, in CDCl_3 , TMS as internal standard). The ^1H NMR spectrum of **1** displayed two aromatic protons at 7.50 ppm appearing as a doublet (H-3'', H-7'') and three other aromatic protons (7.55 ppm; H-4'', H-5'', H-6'') as a multiplet. Moreover, two methyl signals (H-15, H-1') were visible at 0.95 ppm (triplet, 7.0 Hz) and 2.96 ppm (singlet). The tertiary amine structure of **1** was further confirmed by ^1H methylene signals at 4.52 ppm (H-1'') and 3.25 ppm (H-1, triplet, 7.4 Hz). The $^1\text{H}, ^1\text{H}$ -COSY coupling pattern suggested that H-1 and H-15 constituted the beginning and the end, respectively, of an aliphatic chain. In the ^{13}C NMR spectrum four aromatic carbon signals were visible. According to the ^1H and HSQC spectra three of these carbon signals (128.8, 132.9 and 130.2 ppm) are coupled to five protons (7.50 and 7.55 ppm) suggesting the occurrence of a singly substituted benzene ring. Most of the 15 carbon atoms of the aliphatic chain were discernible in the ^{13}C NMR spectrum with the exception of 28.9 ppm signal representing four carbons. Additional evidence for the structure of **1** was furnished by the C,H long-range coupled NMR spectrum (HMBC). Thus $^3J_{\text{C,H}}$ couplings of the *N*-methyl protons (H-1') with C-1 and C-1'', and of H-1'' with C-2'', C-3'' and C-7'' were visible. The structure of **1** thus appears as a hitherto unknown fungal tertiary amine.

Fig. 1. Structure of piptamine (**1**).

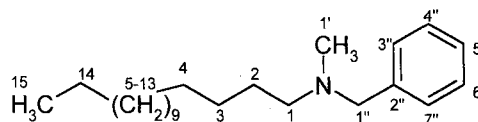


Table 1. Antimicrobial activity of piptamine 1.

Test organisms	MIC [$\mu\text{g/ml}$]
<i>Staphylococcus aureus</i> 134/94	6.25*
<i>Staphylococcus aureus</i> SG 511	0.78*
<i>Enterococcus faecalis</i> 1528	1.56*
<i>Bacillus subtilis</i> ATCC 6633	1.00*
<i>Escherichia coli</i> SG 458	12.5*
<i>Rhodotorula rubra</i> IMET 25030	50.0**
<i>Kluyveromyces marxianus</i> IMET 25148	6.25**
<i>Candida albicans</i> BMSY 212	6.25**
<i>Sporobolomyces salmonicolor</i> SBUG 549	6.25***
<i>Penicillium notatum</i> JP 36	> 50.0 ***

* MIC obtained by NCCLS-method¹⁾

** MIC obtained by NCCLS-method²⁾

*** MIC obtained from agar well diffusion assay³⁾

Piptamine showed antimicrobial activity against a series of Gram-positive bacteria, yeasts and fungi (Table 1). Particularly notable MIC values for *Staphylococcus aureus* SG 511 and *Enterococcus faecalis* 1528, 0.78 $\mu\text{g/ml}$ and 1.56 $\mu\text{g/ml}$, respectively, were obtained¹⁻³⁾. Hemolytic activity of **1** was found at 10~50 $\mu\text{g/ml}$ using heparinized blood of Beagle dogs⁴⁾.

References

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